

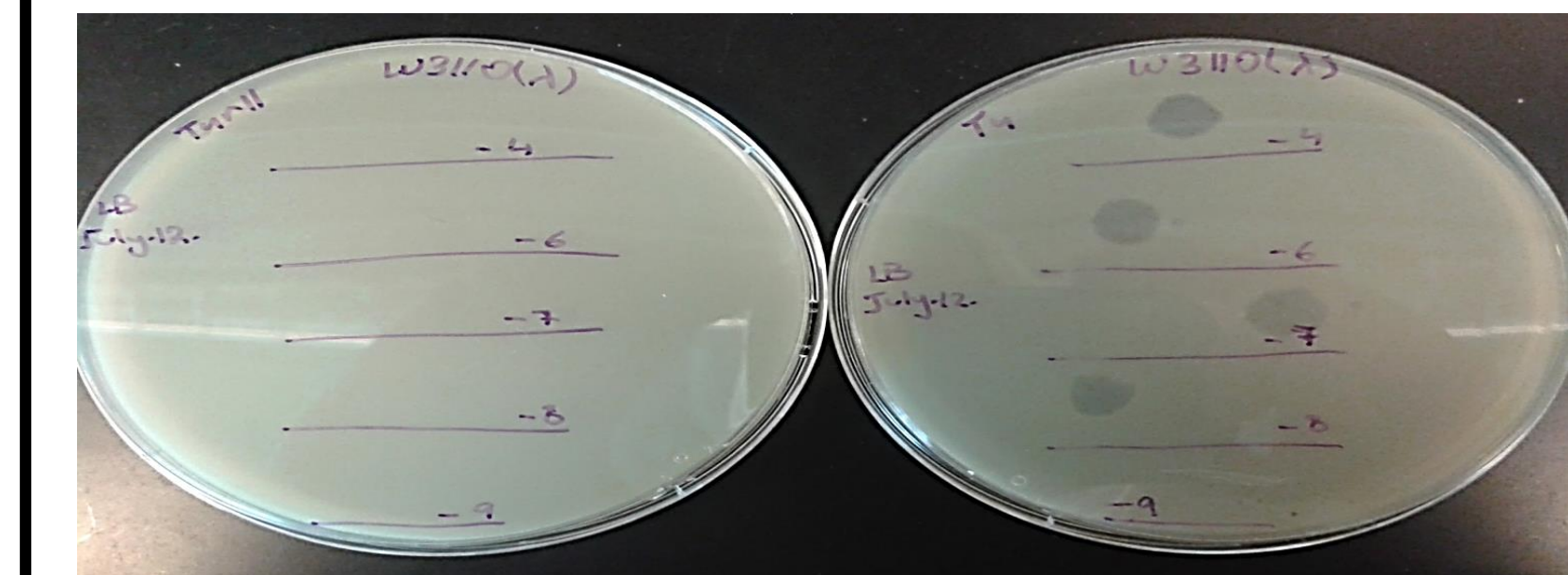
Isolation and Characterization of Host Mutations that Suppress the Bacteriophage Lambda (λ) Rex Phenotype



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Introduction

the λ T4rII exclusion (Rex) phenotype: is the ability of the *rex* genes (*rexA*, *rexB*) of bacteriophage Lambda (λ) to prevent mutant bacteriophage T4rII plaque formation on *E. coli* hosts lysogenized by λ .



More than six decades have now passed since the discovery of Rex by Seymour Benzer in 1955, yet the mechanism behind this elusive exclusion system remains a mystery.

The Rex system is encoded by two genes of λ (*rexA*, and *rexB*); the expression of which is primarily regulated by the repressor gene *cl* from the P_M promoter.

Factors affect the onset and/or abrogation of Rex:

- 1- T4rII infection: results in rapid membrane depolarization and a harsh cellular environment that in many ways resembles stationary phase metabolism and morphology.
- 2- Disruption of the RexA:RexB balance: can lead to same manifestations without infection.
- 3- Ionic environment: Monovalent cations like H^+ , Na^+ in the cellular environment are essential for the exclusionary phenotype. Divalent cations such as Ca^{2+} , Mg^{2+} , or polyamines in culture can attenuate the exclusion activity.

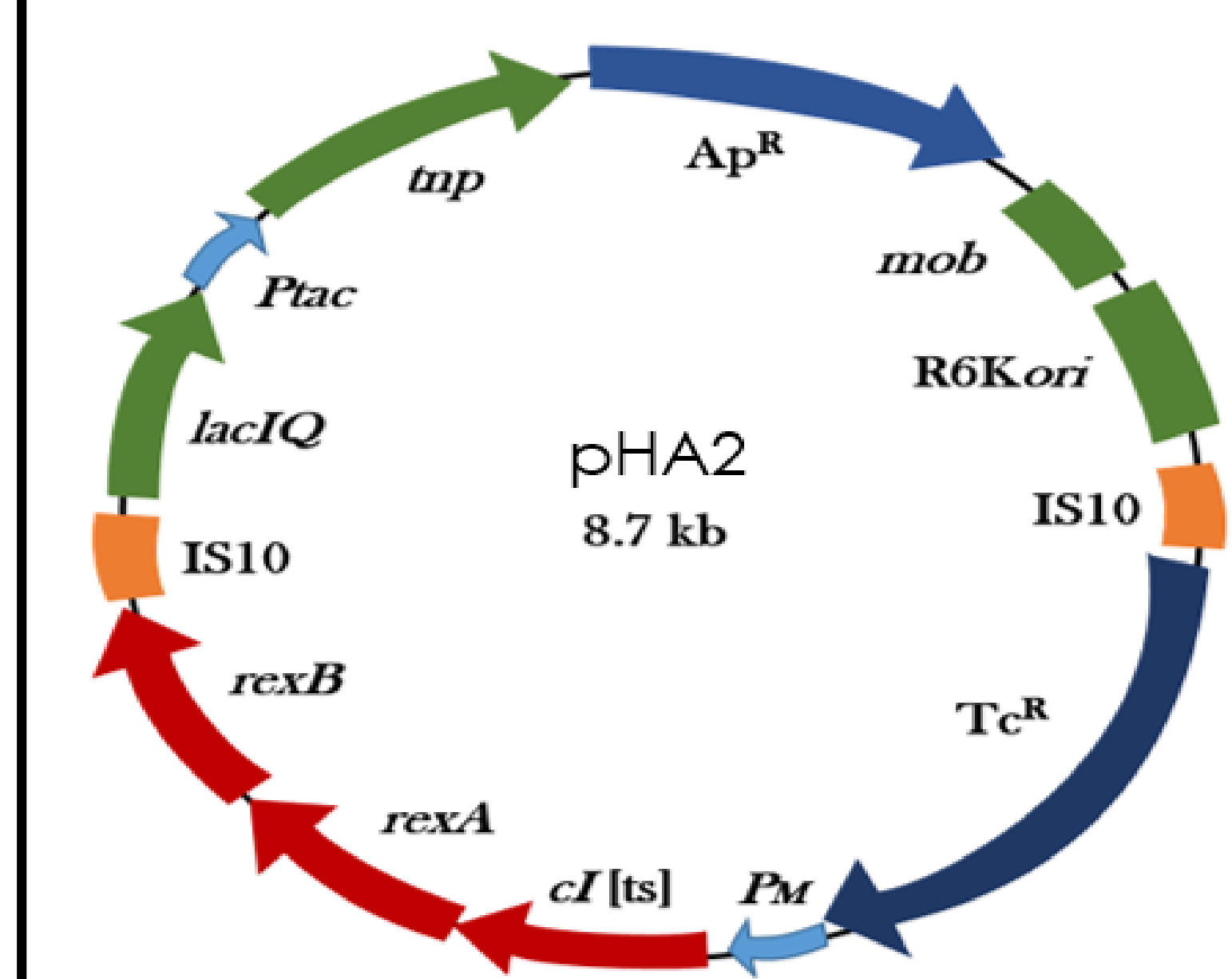
Despite some cell killing, infected lysogens can recover from Rex activation. Rex may thus be a mutualistic protection mechanism that protects both itself and the host cell from external infection.

It has long been believed that additional host proteins like the Outer Membrane Proteins (Omp) are involved in such a pleiotropic phenotype.

We aimed to generate, isolate and characterize *E. coli* mutants that attenuate Rex to uncover important clues about the functionality of Rex and the mechanism of its activation.

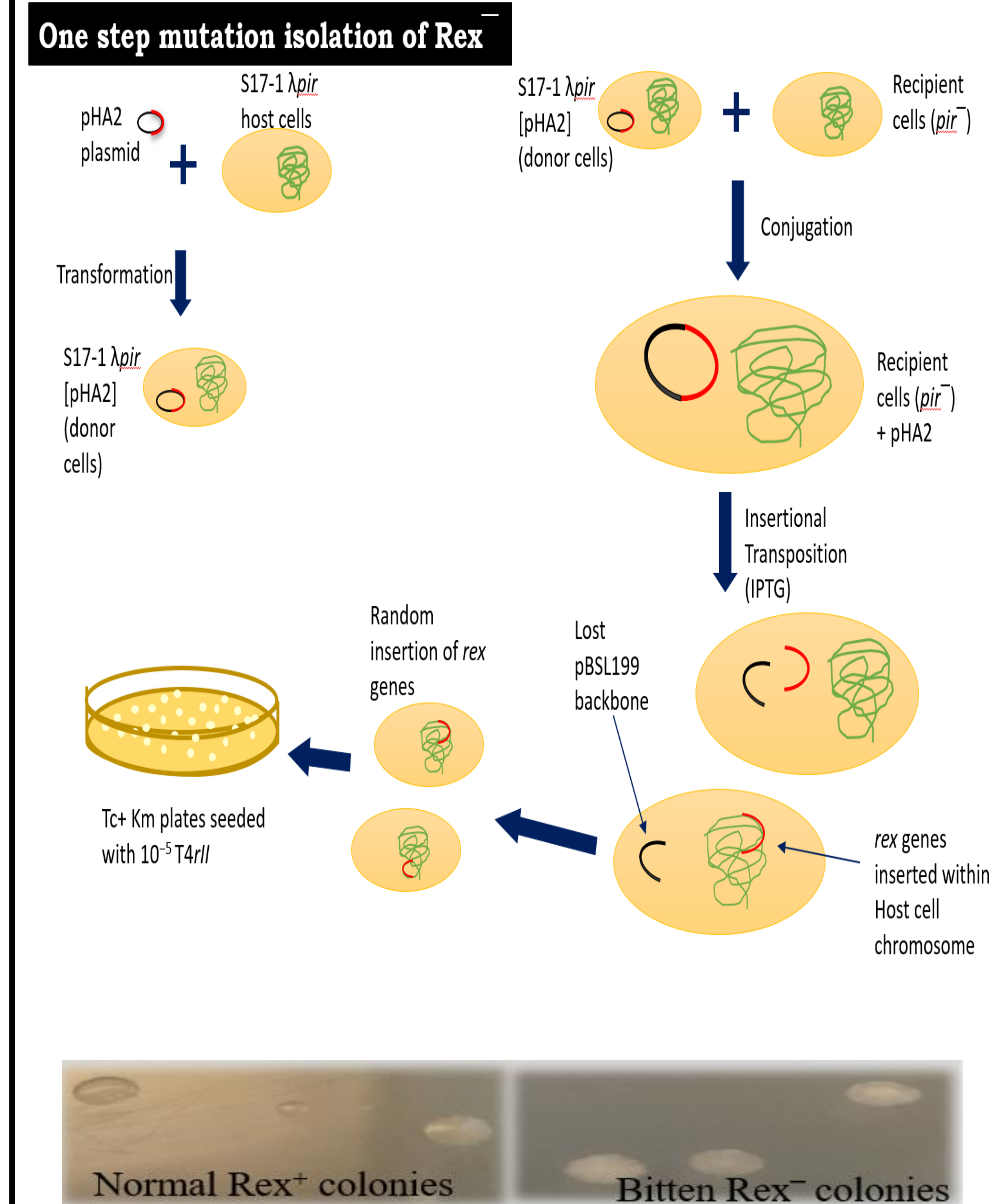
Methods

We have designed a system for the rapid one-step isolation of host mutations that abrogate Rex in order to identify the host genes involved and to elucidate the mechanism of this enigmatic exclusion system.



Construction of the suicide *rex-* vector (pHA2): R6K *ori*: origin of replication which needs π replication protein (*pir* gene) to initiate plasmid replication. *Tn10*: transposable element includes two IS10 insertion sequences flanking tetracycline resistance marker (Tc^R) and the transposase gene. *cl-rexA-rexB*: gene of interest conferring natural *rexA-rexB* expression scenario(*rex-*)

Mutagenize and isolate *rex+* *E. coli* mutants that impact Rex activity:



Results

- We have isolated 13 host mutations of *E. coli* K-12 that attenuated the Rex phenotype as well as identifying for the first time several outer membrane protein genes including: *ompA*, *ompF*, *ompW*, and *ompX* that are involved in imparting Rex activity.
- Confirming isolated *rex+* *E. coli* mutant(s) for sensitivity to T4rII plating:

<i>E. coli</i> # of Mutant	Mutations ($\Delta clpP$) (<i>rex+</i>)	Relative Efficiency of Plating (T4rII)	Rex Activity
JW0427-1	<i>rex-</i>	1.0	$< 1 \times 10^{-7}$
JW0427-1 (λ)	$\lambda^+(rex^+)$	$< 1 \times 10^{-7}$	1.0
JW0427-1	<i>rex+</i>	$< 1.0 \times 10^{-7}$	1.0
# JW-HA 1	<i>rex+</i>	2.3×10^{-5}	< 100 fold
# JW-HA 2	<i>rex+</i>	7.0×10^{-6}	$< \sim 100$ fold
# JW-HA 3	<i>rex+</i>	1.5×10^{-5}	< 100 fold
# JW-HA 4	<i>rex+</i>	1.0×10^{-3}	$< 10K$ fold
# JW-HA 5	<i>rex+</i>	1.7×10^{-5}	< 100 fold
# JW-HA 6	<i>rex+</i>	1.4×10^{-4}	$< 1K$ fold
# JW-HA 9	<i>rex+</i>	5.0×10^{-6}	< 10 fold
# JW-HA 11	<i>rex+</i>	1.6×10^{-5}	< 100 fold
# JW-HA 12	<i>rex+</i>	1.3×10^{-5}	< 100 fold
# JW-HA 19	<i>rex+</i>	2.0×10^{-4}	$< 1K$ fold
# JW-HA 20	<i>rex+</i>	8.0×10^{-6}	$< \sim 100$ fold
# JW-HA 21	<i>rex+</i>	1.6×10^{-5}	< 100 fold
# JW-HA 25	<i>rex+</i>	1.8×10^{-5}	< 100 fold

1-T4/T4rII plating efficiency on Δomp (*rex+*) infective centers:

Test of viable candidate genes postulated to influence Rex activity:

<i>E. coli</i> Strain	Δomp location	Plasmid	Efficiency of Plating		
			T4(wt)	T4rII	Rex
BW25113	wt	pHA1*	0.33	$< 3.0 \times 10^{-7}$	1.0
JW0554-1	$\Delta ompT$	pHA1	0.16	$< 2.0 \times 10^{-7}$	1.0
JW0799-1	$\Delta ompX$	pHA1	0.2	3.0×10^{-4}	$< 10^3$
JW0912-1	$\Delta ompF$	pHA1	0.15	3.0×10^{-5}	$< 10^2$
JW0940-6	$\Delta ompA$	pHA1	3.0×10^{-4}	$< 4.0 \times 10^{-7}$	1.0
JW1248-2	$\Delta ompW$	pHA1	0.22	1.0×10^{-4}	$< 10^3$
JW1312-1	$\Delta ompG$	pHA1	0.12	$< 2.0 \times 10^{-7}$	1.0
JW1371-5	$\Delta ompN$	pHA1	0.3	$< 4.0 \times 10^{-7}$	1.0
JW2203-1	$\Delta ompC$	pHA1	$< 1 \times 10^{-7}$	$< 1.0 \times 10^{-7}$	**
JW3368-1	$\Delta ompR$	pHA1	3.0×10^{-5}	$< 4.0 \times 10^{-7}$	1.0
JW3846-1	$\Delta ompL$	pHA1	0.13	1.7×10^{-5}	$< 10^2$

* PHA1 is pUC19 (*rex+*) plasmid
 ** $\Delta ompC$ mutant was not able to form plaques as OmpC is the absorption site for T4 phage, so neither phage can infect this strain.

2- T4rII plating efficiency on $\Delta omp \lambda cl-857$ lysogens following the infective center assay:

<i>E. coli</i> Strain	Δomp (del) Location	Relative Efficiency of Plating T4rII	Rex Activity
BW25113(wt)	-	$< 3.0 \times 10^{-7}$	1.0
JW0554-1	$\Delta ompT$	$< 2.0 \times 10^{-7}$	1.0
JW0799-1	$\Delta ompX$	4.5×10^{-4}	$< 10^3$
JW0912-1	$\Delta ompF$	3.3×10^{-5}	$< 10^2$
JW0940-6	$\Delta ompA$	5.0×10^{-4}	$< 10^3$
JW1248-2	$\Delta ompW$	1.0×10^{-3}	$< 10^4$
JW1312-1	$\Delta ompG$	$< 2.0 \times 10^{-7}$	1.0
JW1371-5	$\Delta ompN$	$< 4.0 \times 10^{-7}$	1.0
JW2203-1	$\Delta ompC$	$< 1.0 \times 10^{-7}$	1.0
JW3368-1	$\Delta ompR$	$< 4.0 \times 10^{-7}$	1.0
JW3846-1	$\Delta ompL$	3.0×10^{-6}	< 10

3- Cell viability of T4rII-infected Δomp (*rex+*) [pHA1] hosts:

<i>E. coli</i> Strain	Δomp Mutation Location	Plasmid	Relative Efficiency of Plating (T4rII)	Reduction in Cell Viability
BW25113	-	(<i>rex+</i>)	8×10^{-4}	1.0
WT	-	(<i>rex-</i>)	$< 2 \times 10^{-7}$	$< 10^3$
BW25113	-	No plasmid	$< 2 \times 10^{-7}$	$< 10^3$
JW0554-1	$\Delta ompT$	(<i>rex+</i>)	4×10^{-4}	1.0
JW0799-1	$\Delta ompX$	(<i>rex+</i>)	1.2×10^{-6}	$< 10^2$
JW0912-1	$\Delta ompF$	(<i>rex+</i>)	1×10^{-6}	$< 10^2$
JW0940-6	$\Delta ompA$	(<i>rex+</i>)	5×10^{-6}	$< 10^2$
JW1248-2	$\Delta ompW$	(<i>rex+</i>)	7×10^{-6}	$< 10^2$
JW1312-1	$\Delta ompG$	(<i>rex+</i>)	9×10^{-3}	~ 1.0
JW1371-5	$\Delta ompN$	(<i>rex+</i>)	5×10^{-4}	1.0
JW2203-1	$\Delta ompC$	(<i>rex+</i>)	≈ 1	~ 0
JW3368-1	$\Delta ompR$	(<i>rex+</i>)	7×10^{-3}	~ 1.0
JW3846-1	$\Delta ompL$	(<i>rex+</i>)	9×10^{-4}	~ 1.0

Involvement of *E. coli* *omp* mutants in Rex activity:

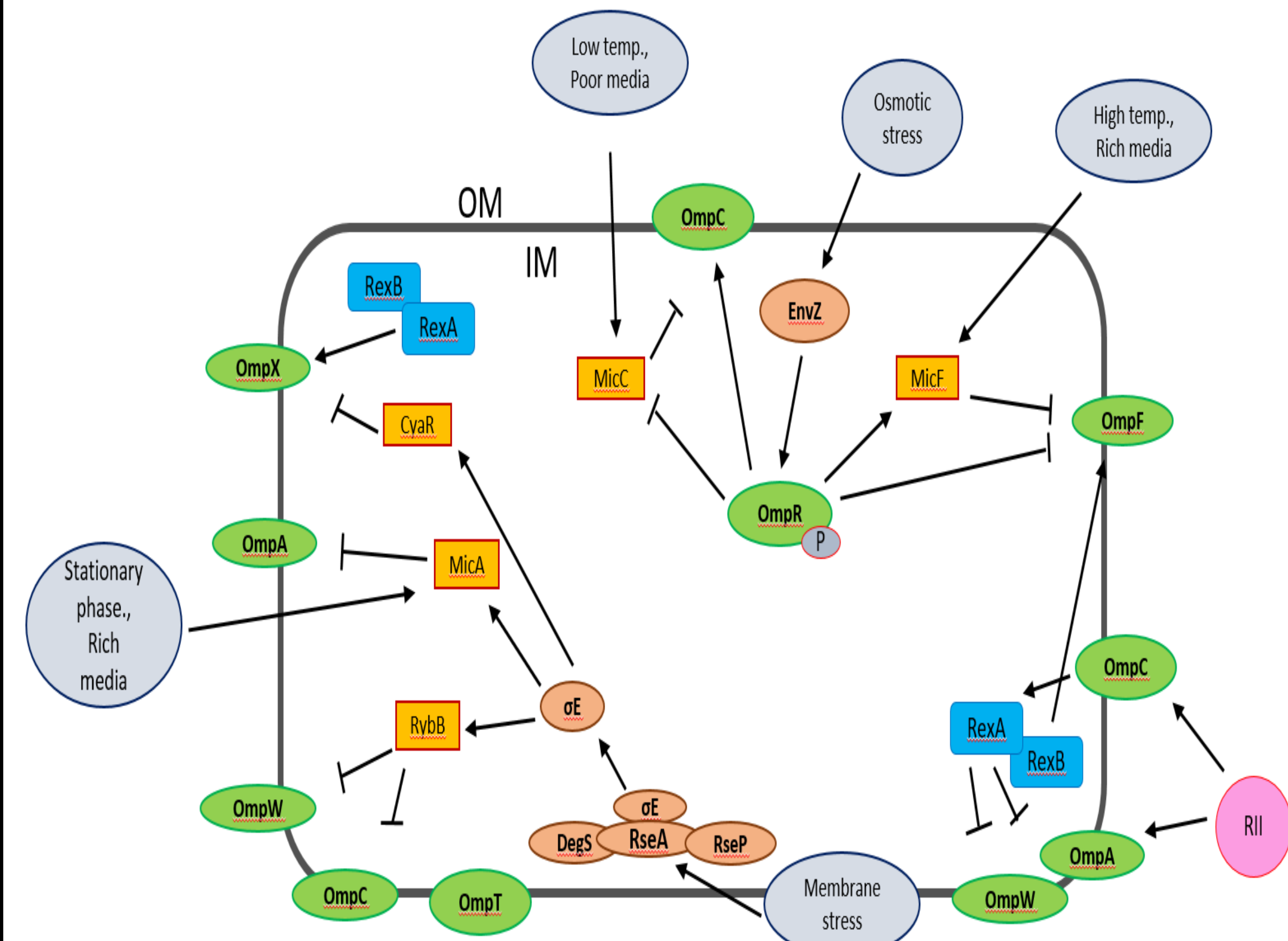


Figure keys: Green ovals are Outer membrane proteins; Pink circles are RII protein of T4 phage; Blue boxes are Rex proteins of λ ; yellow boxes are small RNAs that regulate the transcription of *omp* genes and the expression of Omp proteins; blunted arrows indicate inhibition/downregulation/blocking expression; regular arrows indicate stimulation/upregulation/initiation expression; OM: outer membrane; IM: inner membrane.

Conclusion and significance of this work

- This project aims to understand the mechanism behind the λ Rex phenotype through identifying and understanding the functions of non-essential host proteins that are involved in Rex activity
- We have shown for the first time that OmpA, OmpW, OmpX and OmpF proteins are involved in Rex activity, supporting the model, whereby Rex activation shunts cells into a stationary phase-like state to prevent T4rII growth
- Understanding the mechanism and the functions of *omp* genes that are involved in the suppression of Rex activity is critical to understanding the function of the *rex* genes and the exclusion mechanism
- Identifying the isolated gene that were knocked-out and attenuated Rex phenotype, we definitely can get noteworthy clues about *rex* genes functions and their mechanism of exclusion
- Using the Tandem affinity purification technique to tag Rex and RII proteins and study their interactions and then to identify and localize the Rex proteins in *E. coli* (λ lysogens)
- Understanding Rex can provide important clues and strategies for use toward eukaryotic exclusion systems and control mechanisms

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