



Expression of Cdc25c in *Piper betle* treated drug resistant human colon cancer cells, HT-29

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RESEARCH BACKGROUND

Colon Cancer

- It is the third most common cancer among men and second most common in women (Torre et. al. 2015)
- About 1.4 million cases in total and 693,900 of them resulting in death only in year 2012 (Torre et. al. 2015).
- The existing treatment measures include 5-fluorouracil based chemotherapy (Shakibaei et. al. 2014).

Piper betle

- Piper betle* has shown its cytotoxic effect on HT-29 cells (Ng et. al. 2014).
- Piper betle* locally known as daun sirih, belongs to the family of Piperaceae (Rai et. al. 2011).
- It is one of the targeted cultivated plant under The Bioeconomy Malaysia 2015 (Mohd 2015)
- The phytochemicals found in *Piper betle* includes chavibetol, chavicol, hydroxychavicol, estragole, eugenol, methyl eugenol, hydroxycatechol, caryophyllene, eugenol methyl ether, cadinene, γ -lactone, allyl catechol, p-cymene, cepharadione A, dotriacontanoic acid, tritriacontane, p-cymene, terpinene, eucalyptol, carvacrol, sesquiterpenes, cadinene, caryophyllene, dotriacontanoic acid, hentriacontane, pentatriacontane, stearic acid, n-triacontanol, triotnacontane, piperlonguminine, allylpyrocatechol diacetate, isoeugenol, 1, 8-cineol, α -pinene, β -pinene, sitosterol, β -sitosteryl palmitate, γ -sitosterol, stigmasterol, ursolic acid, and ursolic acid 3 β -acetate (Rai et. al. 2011)
- Its properties include anti-bacterial, anti-cariogenic, anti-fungal, anti-allergic, anti-diabetic, anti-inflammatory, hepatoprotective, cardioprotective, and immunomodulatory effects (Periyanayagam et. al. 2012).
- Past studies showed HT-29 cells appear as round shaped cells after treated with IC₅₀ of *Piper betle* (Gnapragasan 2015; Hay 2015). Round shaped cells may indicate M-phase cells (Sakaue-Sawano et al 2007) that probably undergone mitotic arrest which subsequently leads to cell death (Elliott & Elliott 2009)

Cdc25c

- Cell cycle consists of four phases namely G₁, S, G₂, and M phase.
- To avoid genetic abnormalities, progression through the phases are subjected to controls and checkpoints. At the checkpoints, if the safety requirements are not met, the cycle will be halted.
- A complex between Cdk1 and cyclinB1 plays an important role in regulation of G₂-M transition (Figure 1).
- Dephosphorylation of Cdk1 and hence activation of the Cdk1/cyclinB1 kinase is catalyzed by Cdc25c (Elliott & Elliott 2009). DNA damage may cause activation of Chk1 (Figure 1) which in turn phosphorylates and inactivates Cdc25C (DiPaola 2002). This will inhibit the Cdk 1 thus causing G₂-M phase arrest (DiPaola 2002).
- Cdc25C as shown in previous study involved in Withaferin A (WA) induced G₂-M phase cell cycle arrest. Result showed that the decreased level of Cdc25C protein which is likely to inhibit the activity of the Cdk1/cyclinB1 kinase complex caused the cell cycle arrest of WA-mediated G₂-M (Stan et. al. 2008).

Paclitaxel

- Paclitaxel serves as a positive control in this study.
- Paclitaxel belongs to the taxane family of antitumor drugs.
- Paclitaxel is the first natural product that has been proven to stabilize microtubules (Abal et. al. 2003).
- By promoting the assembly of microtubules, paclitaxel obstructs their depolymerisation thus blocking the progression of cell cycle at the G₂-M phase (Toiyama et. al. 2009).
- Study by Toiyama et. al. (2009) showed HT-29 cells treated with Paclitaxel at cytotoxic doses presented with a significant accumulation of cells in G₂-M phase.

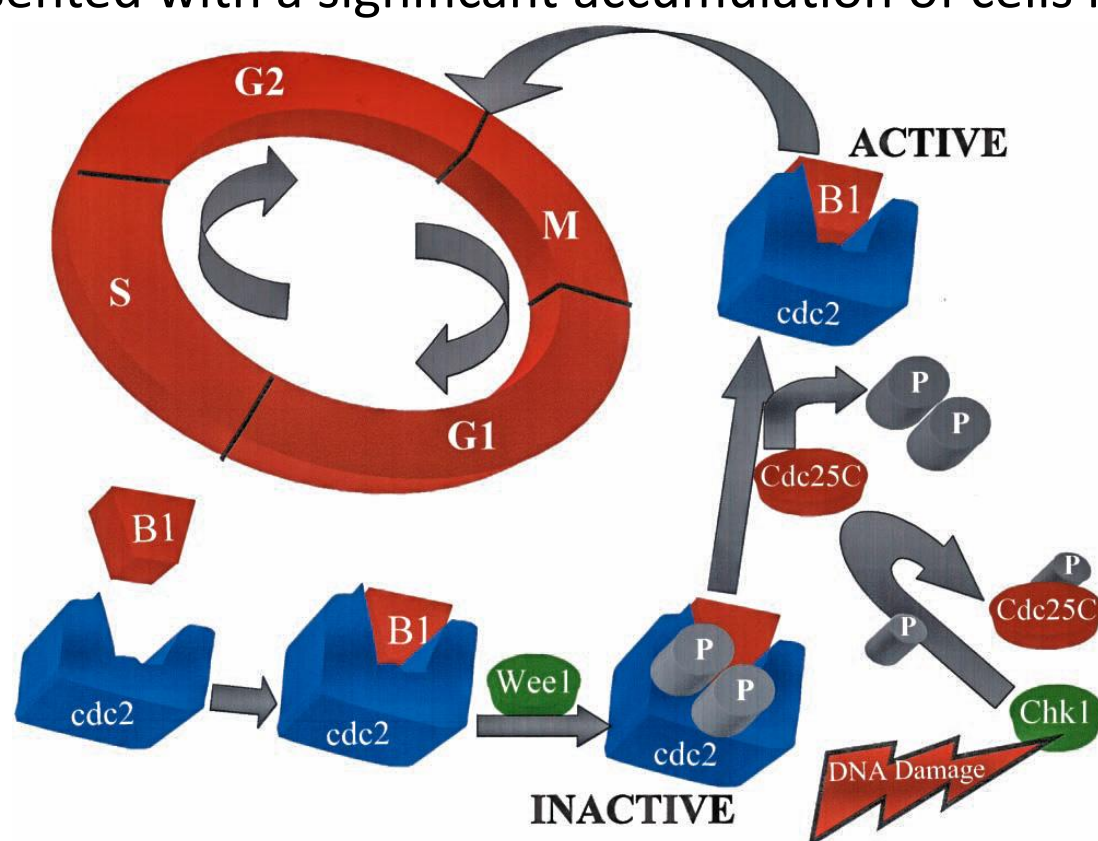
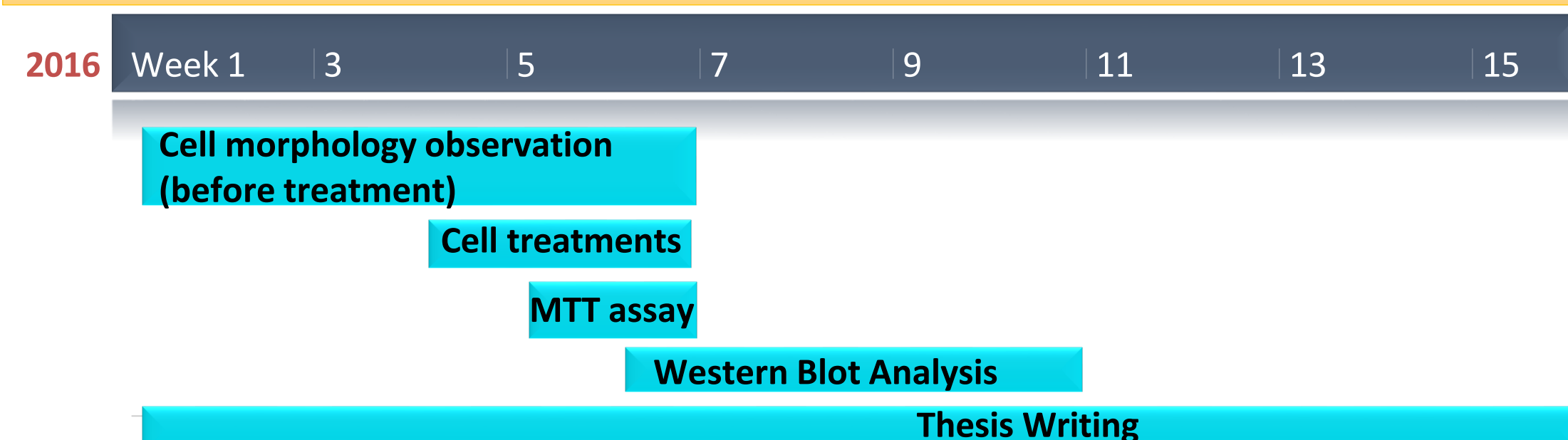


Figure 1: The late G₂ checkpoint controlling cell-cycle progression from G₂ to M phase. (DiPaola 2002).

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RESEARCH QUESTIONS

What is the modulation effect of *Piper betle* on cell cycle that subsequently leads to cancer cell death?

OBJECTIVES

To determine the expression of Cdc25c in *Piper betle* treated drug resistant colon cancer cells.

HYPOTHESIS/ EXPECTED OUTCOME

Piper betle treated HT-29 cells express high abundance of M-phase regulatory protein compared to untreated cells.

METHODOLOGY

HT-29 colon cancer cells (American Type Cell Culture) will be cultured in McCoy's 5A medium + 10% (v/v) Fetal Bovine Serum and incubated in the condition of 37°C and 5% CO₂.

Morphology of the cells before treatment will be observed under inverted microscope.

| Control HT-29 (without treatment) | HT-29 cells treated with Paclitaxel | HT-29 cells treated with <i>Piper betle</i> extract |
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Morphology of the cells after treatment will be observed under inverted microscope.

MTT assay: HT-29 cells treated with Paclitaxel will be plated at 10,000 cells per well and incubated for 6 to 24 hours. Ten (10) μ L yellow tetrazolium MTT (3-(4, 5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide) will be added and incubated for 2 to 4 hours until purple precipitate is visible. 100 μ L Detergent Reagent will be added and left in the dark for 2 hours. The absorbance is read at 570nm to determine the cell viability.

Western blot (Figure 2): Cells will be lysed using a lysis buffer and centrifuged. The supernatant will be collected and protein concentrations will be determined by the Lowry Protein Assay. Samples will then be separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) using a 10% gel, and transferred to a polyvinylidene fluoride (PVDF) membrane using the Trans-Blot Turbo Transfer System (Bio-Rad). Blocking agent will be used to block unspecific sites. Primary and secondary antibodies will be added. Visualization by chemiluminescence will be done (Figure 2).

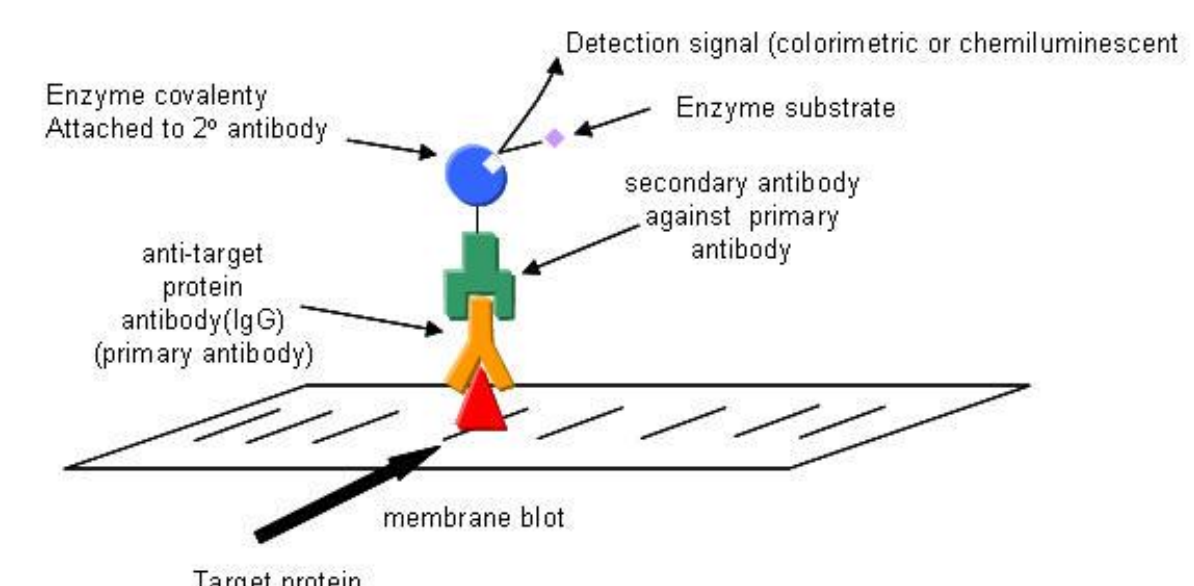


Figure 2: The schematic diagram of the principles of western blotting (College of Saint Benedict & Saint John's University 2006).

REFERENCES

- Abal, M, Andreu, J & Barasoain, I 2003, 'Taxanes: microtubule and centrosome targets, and cell cycle dependent mechanisms of action.', *Curr Cancer Drug Targets*, vol. 3, no. 3, pp. 193-203.
- College of Saint Benedict & Saint John's University 2006, Electrophoresis, <<https://employees.csbsju.edu/hjakubowski/classes/ch331/Techniques/TechElectrophoresis.htm>>.
- DiPaola, R 2002, 'To Arrest or Not To G₂-M Cell-Cycle Arrest', *Clinical Cancer Research*, vol. 8, pp. 3311-4.
- Elliott, W & Elliott, D 2009, *Biochemistry and Molecular Biology*, 4 edn, Oxford University Press Inc., United States.
- Gnapragasan, S 2015, 'Investigation of Tubulin Structure and Network in *Piper betle* Linn. and 5-Fluorouracil Treated Human Colon Cancer Cells, HT-29', BSc Thesis thesis, Taylor's University.
- Hay, Y 2015, 'Investigation of Caspases Activity in *Piper Betle* Linn. and 5-Fluorouracil-Treated Colon Cancer Cells', BSc Thesis thesis, Taylor's University.
- Mohd, N 2015, 'Leveraging Borneo's Strength through Value-driven Bioeconomy', paper presented to Bioborneo 2015 Conference Programme, Grand Ballroom, The Magellan Sutra Resort, Kota Kinabalu, Sabah, <http://bioborneo2015.sedia.com.my/images/papers/P21_BioEconomy_Initiative_Dr_nazleeKamal.pdf>.
- Ng, P, Rajab, NF, Then, SM, Mohd Yusof, Y, Wan Ngah, WZ, Pin, KY, & Looi, ML 2014, 'Piper betle leaf extract enhances the cytotoxicity effect of 5-fluorouracil in inhibiting the growth of HT29 and HCT116 colon cancer cells', *Journal of Zhejiang University-SCIENCE B (Biomedicine & Biotechnology)*, vol. 15, no. 8, pp. 692-700.
- Periyanayagam, K, Jagadeesan, M, Kavimani, S, & Vetriselvan, T 2012, 'Pharmacognostical and Phyto-physicochemical profile of the leaves of *Piper betle* L. var *Pachaikodi* (Piperaceae) - Valuable assessment of its quality', *Asian Pacific Journal of Tropical Biomedicine*, pp. 506-10.
- Rai, M, Thilakchand, K, Palatty, P, Rao, P, Rao, S, Bhat, H, & Baliga, M 2011, 'Piper betel Linn (Betel Vine), the Malignant Southeast Asian Medicinal Plant Possesses Cancer Preventive Effects: Time to Reconsider the Wronged Opinion', *Asian Pacific Journal of Cancer Prevention*, vol. 12.
- Sakaue-Sawano, A, Kurokawa, H, Morimura, T, Hanyu, A, Hama, H, Osawa, H, Kashiwagi, S, Fukami, K, Miyata, T, Miyoshi, H, Imamura, T, Ogawa, M, Masai, H, & Miyawaki, A 2008, 'Visualizing Spatiotemporal Dynamics of Multicellular Cell-Cycle Progression', *Cell*, vol. 132, pp. 487-98.
- Shakibaei, M, Buhrmann, C, Kraehe, P, Shayan, P, Lueders, C, & Goel, A 2014, 'Curcumin Chemosensitizes 5-Fluorouracil Resistant MMR-Deficient Human Colon Cancer Cells in High Density Cultures', *PLoS ONE*, vol. 9, no. 1, pp. 1-12.
- Stan, S, Zeng, Y, & Singh, S 2008, 'Ayurvedic Medicine Constituent Withaferin A Causes G₂ and M Phase Cell Cycle Arrest in Human Breast Cancer Cells', *Nutrition and Cancer*, vol. 60, no. 1, pp. 51-60.
- Toiyama, Y, Inoue, Y, Hiro, J, Ojima, E, Watanabe, H, Narita, Y, Okigami, M, Hosono, A, Miki, C, & Kusunoki, M 2009, 'Paclitaxel inhibits radiation induced VEGF secretion and enhances radiosensitizing effects in human colon cancer cell HT29', *Cancer Therapy*, vol. 7, pp. 123-32.
- Torre, L, Bray, F, Siegel, R, Ferlay, J, Lortet-Tieulent, J & Jemal, A 2015, 'Global Cancer Statistics, 2012', *Cancer Journal for Clinicians*, vol. 65, no. 2, pp. 87-105.