

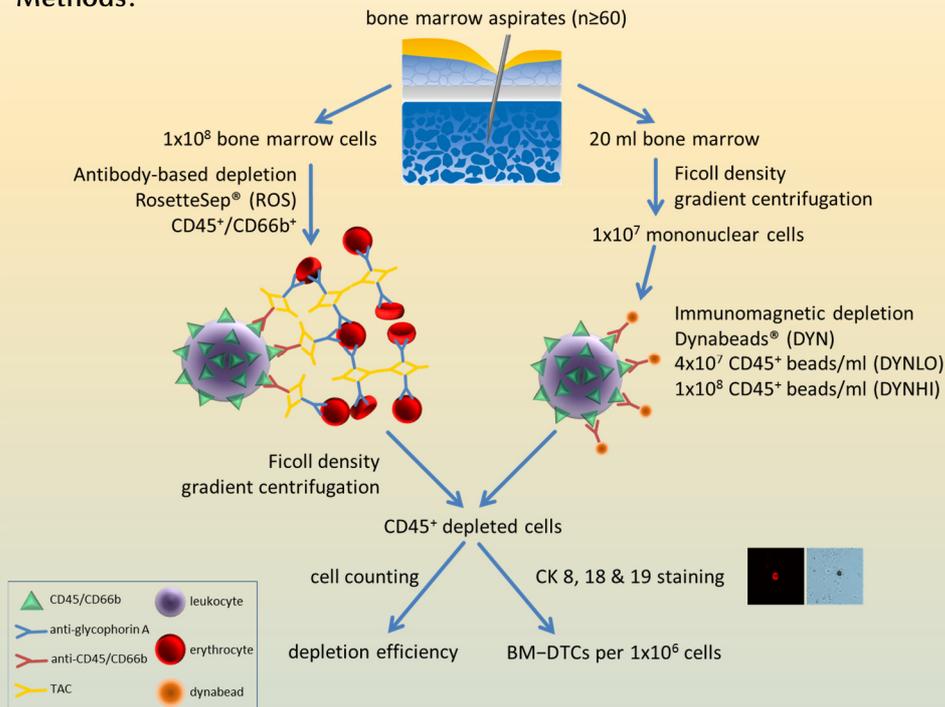
Comparison of CD45⁺ depletion methods for enrichment of disseminated tumor cells in bone marrow samples

Birte Möhlendick^{1*}, Imke Hoffmann¹, Swetlana Seidschner¹, Sarah Schumacher¹, Alexander Smyczek¹, Carina Vaerst¹, Wolfram T. Knoefel¹ and Nikolas H. Stoecklein¹

¹Department for General, Visceral, and Pediatric Surgery, University Hospital of the Heinrich-Heine University Düsseldorf, contact: birte.moehlendick@uni-duesseldorf.de

Background: Disseminated tumor cells (DTC) in bone marrow (BM) samples are, with a frequency of approx. one DTC per 1×10^6 cells, extremely rare cells. The most common method to enrich DTCs is Ficoll density gradient centrifugation. In order to increase the efficiency of bone marrow-DTC enrichment we tested three different strategies to deplete CD45⁺ cells.

Methods:



Results:

- CD45⁺ cells could be depleted most efficiently up to 500-fold using ROS, followed by 44-fold with DYNHI and 20-fold using DYNLO (Fig.1A)
- DTCs could be detected in 32.7% (DYNLO), 12.5% (DYNHI) and 72.1% (ROS) of the patient samples
- The immunomagnetic depleted DYNLO samples (mean=0.48 per 1×10^6 cells) had slightly higher DTC counts (8.33 vs 6.35 DTCs per 1×10^6 cells) compared to the DYNHI samples (mean=0.39 per 1×10^6 cells) (Fig.1B)
- DTC numbers were significantly higher ($p < 0.0001$) in the ROS samples (mean=5.60 per 1×10^6 cells) with up to 40 DTCs per 1×10^6 cells (Fig.1B)

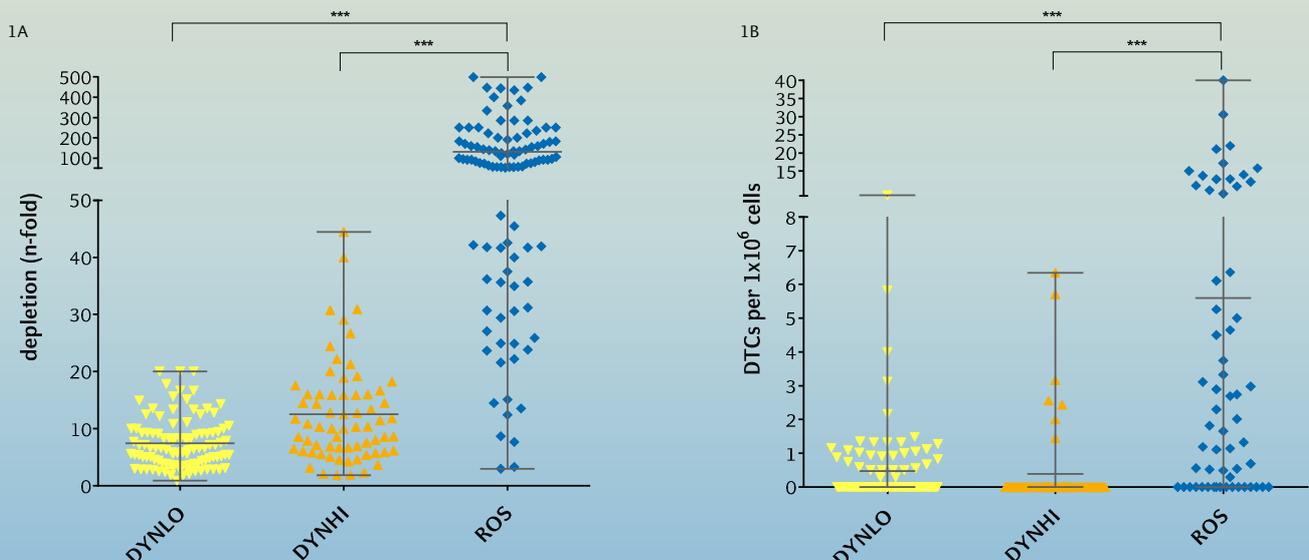


Figure 1: A) Depletion efficiencies of the three different strategies. A significantly higher depletion efficiency was observed in the ROS samples ($p < 0.0001$, Mann-Whitney test) compared to the immunomagnetic depletion strategies DYNLO and DYNHI. **B)** Number of detected DTCs per one million cells in the depleted bone marrow samples. A significantly higher number of DTCs was observed in the ROS samples ($p < 0.0001$, Mann-Whitney test) compared to the immunomagnetic depletion methods DYNLO and DYNHI.

Conclusions: In our hands the ROS procedure was most efficient in depleting CD45⁺ cells from bone marrow samples. This method enabled a high input of bone marrow material (1×10^8 mononuclear bone marrow cells) leading to an increased DTC detection rate in bone marrow samples from tumor patients.